

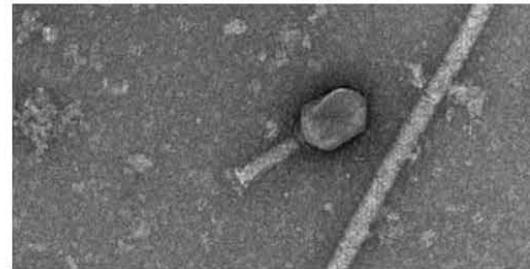
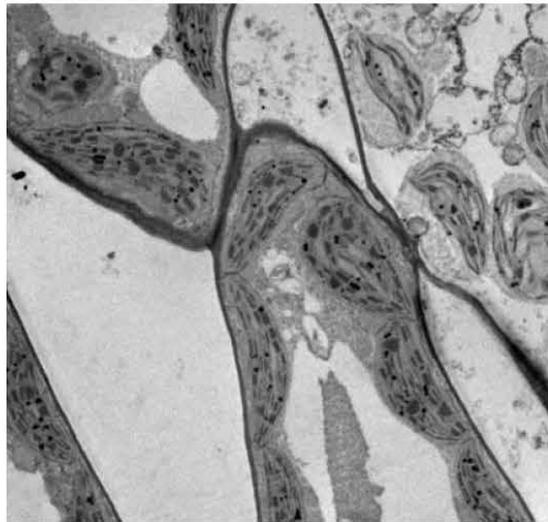
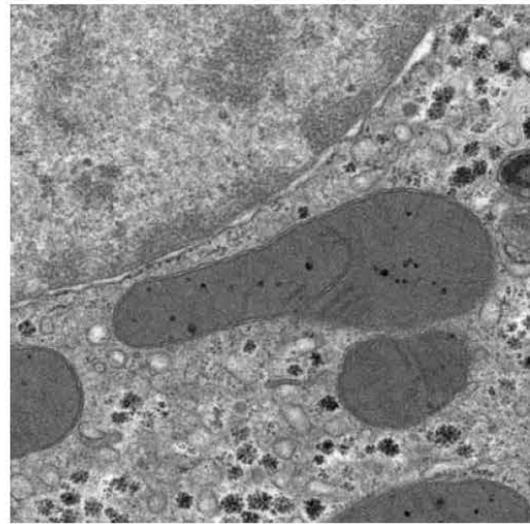


Introducing...

# UransyLess

EM STAIN

A Substitute for Uranyl Acetate



 **DELTA**  
MICROSCOPIES

## A Substitute for Uranyl Acetate...

# UranylLess EM Stain

EMS is proud to introduce UranylLess, a new contrast stain solution for TEM, for all of your negative staining applications. It is an amazing substitute for Uranyl Acetate with similar results.

After only a minute of contact, UranylLess' fast-acting, non-radioactive lanthanide mix is finished staining your sections or deposits (see protocols below). If needed, lead citrate is recommended to increase the contrast.

UranylLess's pH level is about 6,8 to 7. The 30ml airless bottle will stain approximately 1500 grids. The airless bottle increases the shelf life, eliminates CO<sub>2</sub> contamination, and produces less waste — the solution pumps out in perfect amounts without leaking or spilling. UranylLess is also available in a larger amount for use in automated staining equipment. When using UranylLess for automated staining, do not wash longer than 10 minutes or you run the risk of losing all contrast.

UranylLess has been tested on many biological tissue (animal and plant): intestine, skeletal and cardiac muscle, liver, kidney, adrenal gland, nerve, cell culture, plant tissue, and also on negative staining of bacteriophage, bacteria, and polymers. UranylLess is ideal because of its ability to stain any kind of material and results are reproducible.

30ml Airless Bottle

### How does an airless bottle operate?

Its use is very simple; simply push on the head of the bottle to get a drop. When you release, the bottle back pump actuator lifts up. It prevents any air inlet in the bottle.

### What is the advantage of an airless bottle?

It is a bottle in which air never enters. Some products, such as lead citrate, are atmospheric CO<sub>2</sub> sensitive. Thanks to this system, those products have a longer shelf life. It also allows the product to be deposited drop by drop, quickly, cleanly and in any position.



### References

"Easier and Safer Biological Staining: High Contrast UranylLess Staining of TEM Grids"

1. Delta Microscopes, 22, B route de saint Ybars, La côte blanche, 31190, Mauressac, France
2. Université Toulouse, CMEAB Faculté Médecine, 118 route Narbonne, 31062, Toulouse, France
3. Microscopy Innovations LLC, 213 Air Park Rd, Suite 101, Marshfield, WI, 54449, USA

"C-Nap1 mutation affects centriole cohesion and is associated with a Seckel-like syndrome in cattle." Nature Communications, Published 23 Apr 2015. Sandrine Floriot, all.



200ml Bottle

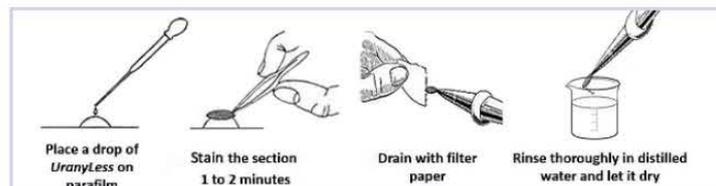
## PROTOCOLS OF USE

### Classic Contrast

This protocol is used for double staining with UranylLess/Lead citrate on ultrathin sections. This protocol is adapted to biologic samples that have been fixed with glutaraldehyde, osmium, or ruthenium and embedded in an epoxy type resin (Epon, Araldite, Spurr) or acrylic type (LRWhite, HM20).

#### Staining Protocol:

- Place a drop of UranylLess on parafilm or any other hydrophobic slide.
- Place the grid on the UranylLess drop for 1 to 2 minutes.
- Blot the grid on a filter paper and then wash in distilled water.
- Let it dry.
- After drying, go to the lead citrate staining according to Reynolds method (1963).
- Place the grid on the lead citrate drop according to the Reynolds method, for 1 minute.
- Blot the grid on a filter paper before rinsing with distilled water.
- Let it dry.



### Technical Tip:

UranylLess is not air or light sensitive, unlike Uranyl Acetate.

After lead citrate, drain immediately in a freshly prepared distilled water bath or wash with 0.01N of NaOH solution.

If there is a precipitate in the solution, filter it prior to use.

If solution was refrigerated, allow solution to return to room temperature prior to use.

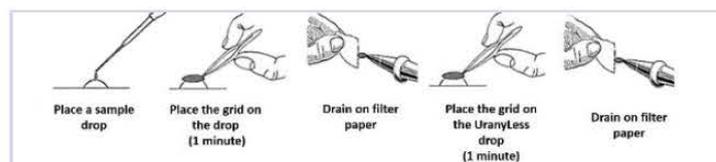
Do not keep lead citrate refrigerated.

### Negative Staining

Negative staining is a very useful technique in electron microscopy. It allows characterization of isolated particles of morphology as bacteria, virus, protein, nanoparticles, liposomes, exosomes, etc.

#### Staining Protocol:

- On a piece of parafilm or any other hydrophobic carrier, place a drop of your solution (~10µl) and a UranylLess drop.
- Using our fine tweezers, place your sample drop on a formvar-carbon coated grid, for about 1 minute.
- Blot your grid using filter paper.
- Place your grid on the UranylLess solution for 1 minute.
- Blot, let it dry for 5 minutes and observe under the microscope.



### Technical Tip:

If the staining is too intense, wash with water for 1 minute.

## Ordering Information

RT	22409	UranylLess EM Stain*	30 ml
RT	22409-20	UranylLess EM Stain	200 ml

\* in airless bottle

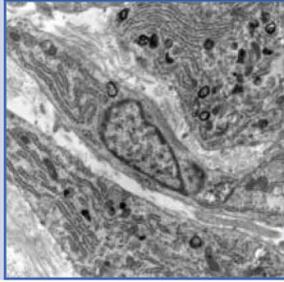




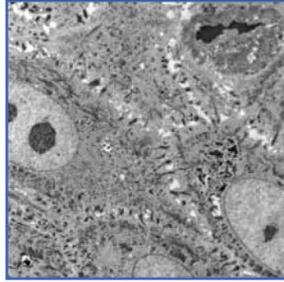
## Reconstituted Epidermis

Preparation of the sample using the following protocol:

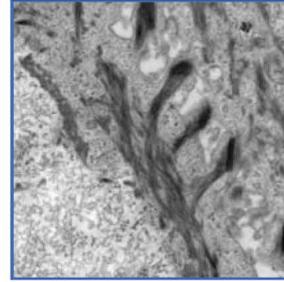
- Fixing Classic Glutaraldehyde, Osmium, Epon / Araldite
- Cutting Ultra-Thin, Double UranylLess Contrast and Lead Citrate



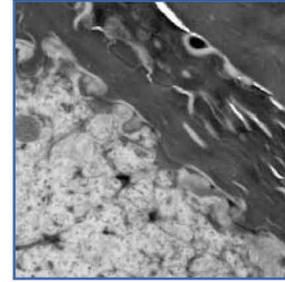
Epidermis. Photo: Audrey Houcine (CMEAB Toulouse)



Epidermis. Photo: Audrey Houcine (CMEAB Toulouse)



Epidermis. Photo: Audrey Houcine (CMEAB Toulouse)

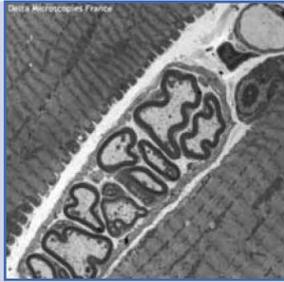


Epidermis. Photo: Audrey Houcine (CMEAB Toulouse)

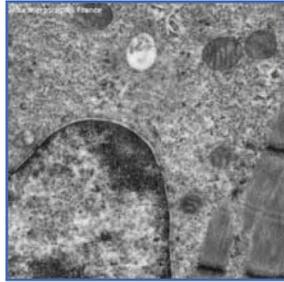
## Muscle - Nerve - Mice

Preparation of the sample using the following protocol:

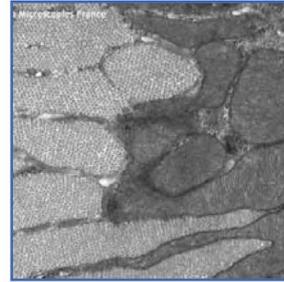
- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - UranylLess Contrast 1 minute followed lead Citrate 1 minute



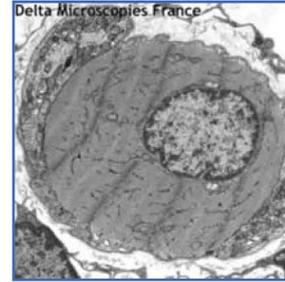
Longitudinal Section of Mouse Skeletal Muscle - Nerve Cup (dense area myelin sheath). Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Detailed View of Myocytes. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Cross Section of Muscle Fibers - Mitochondria. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

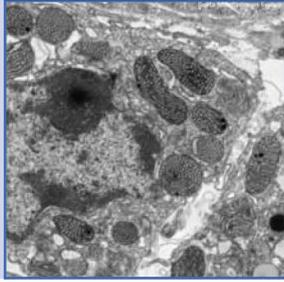


Myocyte. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

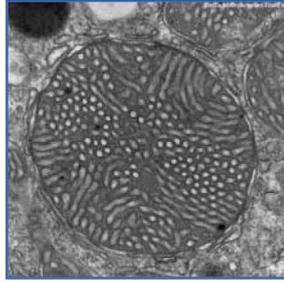
## Mouse Ovarian Follicle

Preparation of the sample using the following protocol:

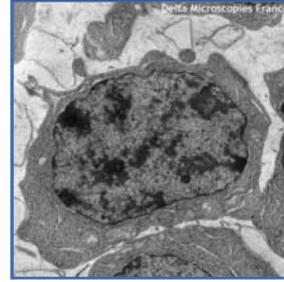
- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UranylLess Lead -Citrate



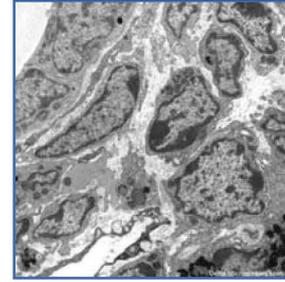
Theca interna Mouse Ovarian Follicle. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Mitochondria in Typical Finger Glove Steroid Synthesis in Cells (Internal Thèque Ovarian Follicle). Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Follicular Cell of the Corona Radiata a Mouse Ovarian Follicle. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

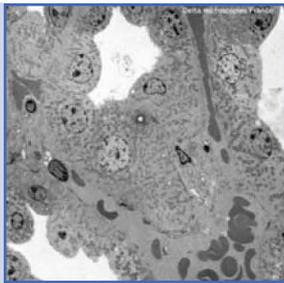


Cell of the External Layer of Mouse Ovarian Follicle. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

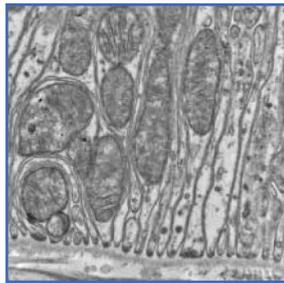
## Mouse Kidney

Preparation of the sample using the following protocol:

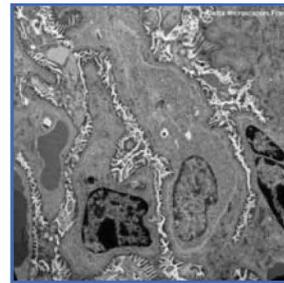
- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - UranylLess Contrast 1 minute followed lead Citrate 1 minute



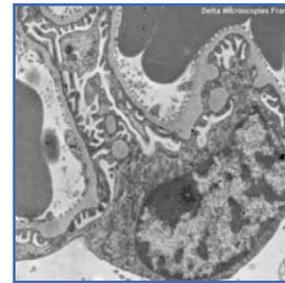
Mouse Kidney. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Basal Invaginations - Hemidesmosome - Basal Lamina: Increase the Exchange Surface - Kidney. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Glomerular area - Podocytes - Stalks - Kidney. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

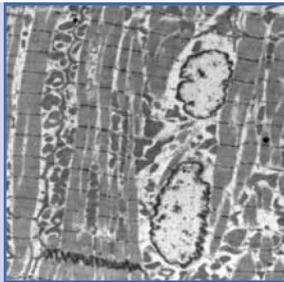


Podocyte - Pedicels - Kidney. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

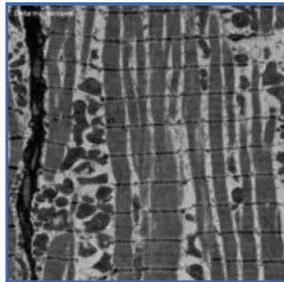
## Mouse Cardiac Muscle

Preparation of the sample using the following protocol:

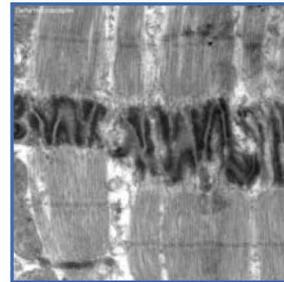
- Classic - Fixing Glutaraldehyde, Osmium, Epon Ultrafine
- Cups, Double UranylLess Contrast and Lead Citrate



Mouse Cardiac Muscle. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Mouse Cardiac Muscle. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

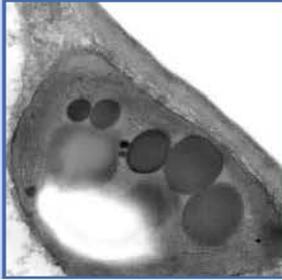


Mouse Cardiac Muscle. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

### Plant Tissue

Preparation of the sample using the following protocol:

- Glutaraldehyde Fixation Classic - Osmium - Included In Epon
- Contrast the UransLess monitoring Lead Citrate



Plant Leaf. Photo: Jeannine Lherminier (INRA - Dijon)

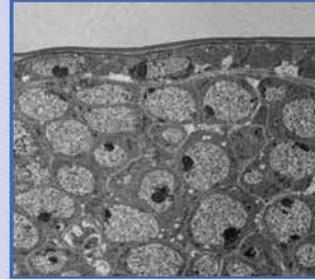


Plant Leaf. Photo: Jeannine Lherminier (INRA - Dijon)

### Drosophila Larva

Preparation of the sample using the following protocol:

- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UransLess lead -citrate

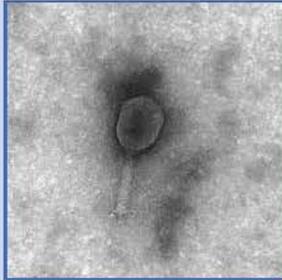


Drosophila Larva. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

### Phage T6

Preparation of the sample using the following protocol:

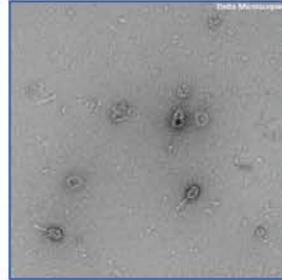
- Staggering Phage T6 on a G300-Cu grid Covered with a Carbon Formvar Film. Ionization 1 minute



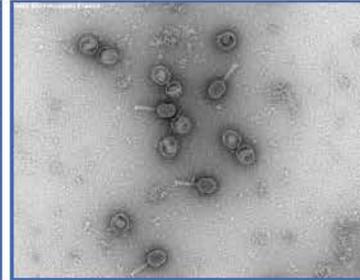
Phage. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Phage. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



Phage. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

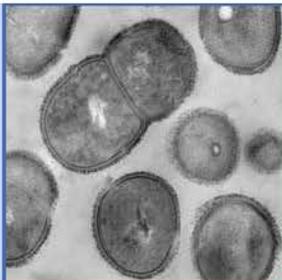


Phage. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

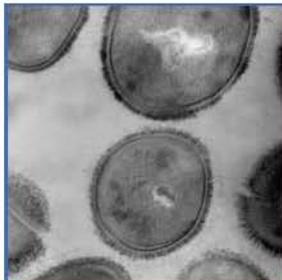
### Cross-Sectional Bacteria

Preparation of the sample using the following protocol:

- Fixing Classic Glutaraldehyde, Osmium, EPON
- Cutting Ultrafine, Double Contrast UransLess and Lead Citrate.



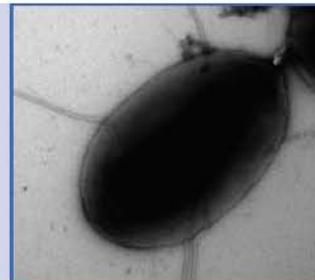
Bacteria. Photo: Christine Longin (INRA Jouy en Josas).



Bacteria. Photo: Christine Longin (INRA Jouy en Josas).

### Bacteria E. Coli

Negative Staining for 2 Minutes UransLess Bacteria Like E. Coli (Adherent and Invasive (ACSI) LF82) Which Have Pili and Flagella.

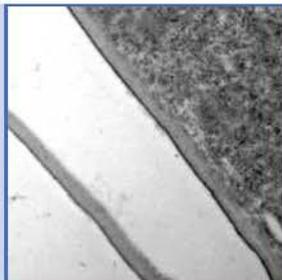


Bacteria. Photo: M2ISH team of Clermont Ferrand

### Sacculina Crustaceans (Small Parasitic Crustacean)

Preparation of the sample using the following protocol:

- Classic Glutaraldehyde Fixation, Osmium, Epoxy Inclusion
- Fine Cups - Contrast to the Aqueous UransLess to 60°C on a Hotplate without Lead Citrate Post Coloring

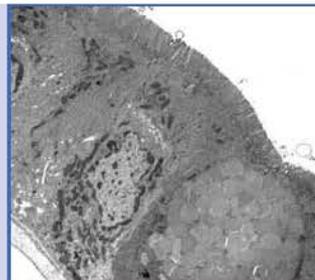


Sacculina (small parasitic crustacean) cuticle area. Photo: Djedjet Chakib (Natural History Museum, Paris)

### Intestine

Preparation of the sample using the following protocol:

- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UransLess Lead -Citrate

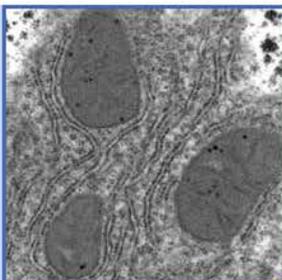


Intestine. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

### Liver Mouse and Gerbil Sahara

Preparation of the sample using the following protocol:

- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UransLess Lead - Citrate

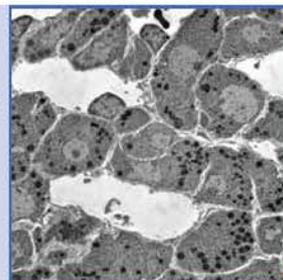


Hepatocyte - Perinuclear Region. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

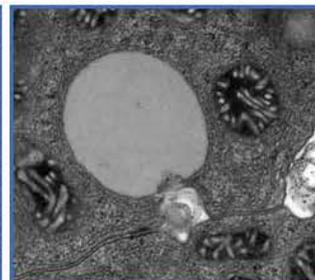
### Adrenal Gland Gerbil Sahara

Preparation of the sample using the following protocol:

- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UransLess lead -citrate



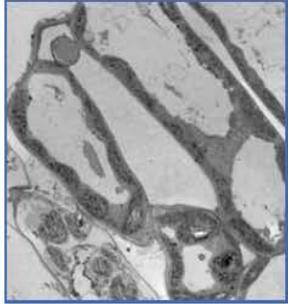
Adrenocortica. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)



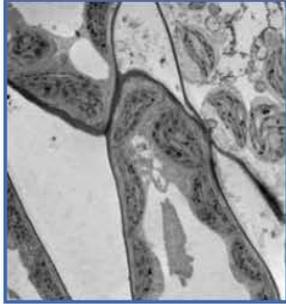
Adrenocortica. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)

**Parsley and Rosebush,** Preparation of the sample using the following protocol:

- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UranylLess lead -citrate



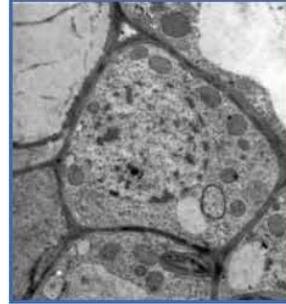
*Parsley Leaf. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



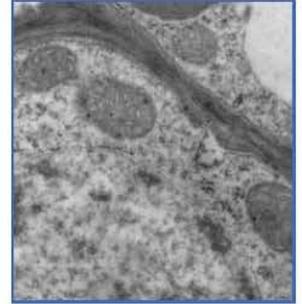
*Parsley Leaf. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



*Rosebush. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



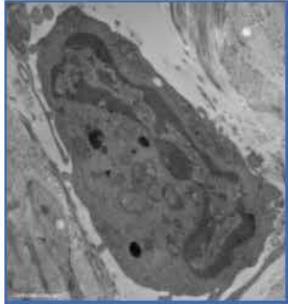
*Rosebush Root. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



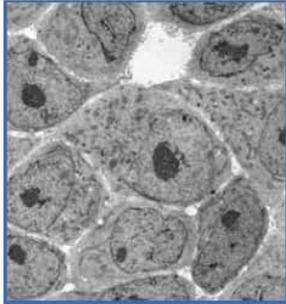
*Rosebush Root. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*

**Culture Cells,** Preparation of the sample using the following protocol:

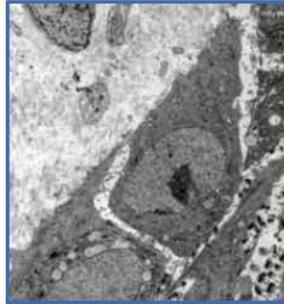
- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - Contrast UranylLess lead -citrate



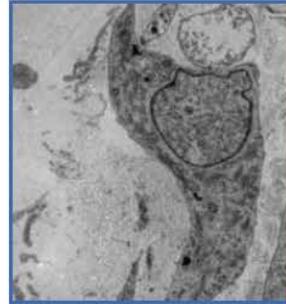
*Culture Cell. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



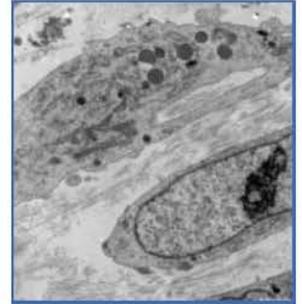
*Culture Cell. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



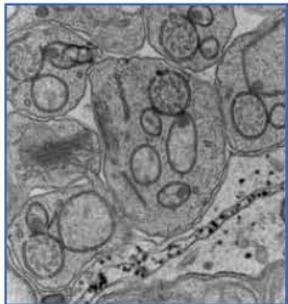
*Culture Cell. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



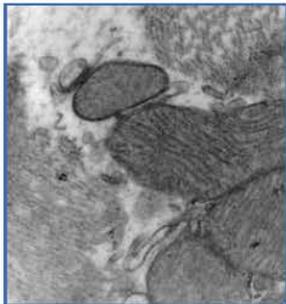
*Culture Cell. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



*Culture Cell. Photo: Nacer Benmeradi (R & D - DeltaMicroscopies-France)*



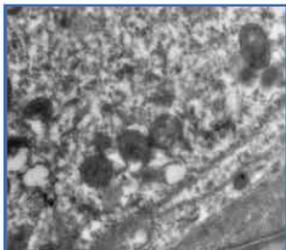
*Spermatid Drosophila. Photo: Chantal Cazevielle Montpellier*



*Heart Headset. Photo: Chantal Cazevielle CRIC / IURC la'INSERM Montpellier (R & D - DeltaMicroscopies-France)*



*Drosophila. Photo: Chantal Cazevielle CRIC / IURC la'INSERM Montpellier*



*Drosophila. Photo: Chantal Cazevielle CRIC / IURC la'INSERM Montpellier*

**PLC Contrast Leica EM Stain**

Preparation of the sample using the following protocol:

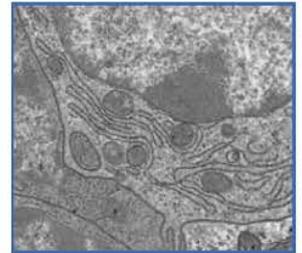
- Classic-Glutaraldehyde Fixation PFA, Osmium, Epon
- Ultrafine Cup - contrast UranylLess Lead -Citrate

Chantal Cazevielle CRIC / IURC INSERM Montpellier tested aqueous UranylLess in the Leica brand grid contrast controller on different tissues, Drosophila heart atrium, retina, cochlea and ileum (Gut). The tissues were fixed according to the standard protocol 2.5% Glutaraldehyde in PHEM buffer, the post fixation in 0.5% osmium in 0.8% potassium ferrocyanide in RT for 2 hours. The sections are collected on single-hole or 200 mesh grids.

The treatment of the grids is UranylLess 7mn lead citrate followed 7 minutes.

We present here some images made by Hitachi transmission electron microscope with a digital camera AMT.

You will notice that the combined action of potassium ferrocyanide and UranylLess reveal a marked way the cyto-membranes in the ileum.



*Drosophila. Photo: Chantal Cazevielle CRIC / IURC la'INSERM Montpellier*